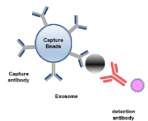




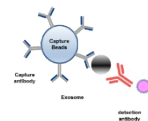
# Flow cytometry methods and kits for the isolation and characterization of EVs

# Our Portfolio

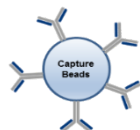
Human Exosome Detection Kits



Mouse Exosome Detection Kits



Capture Beads



Exosome precipitation solution



Lyophilized Exosome Standards



Antibodies



Secondary Antibodies

Exosome Isolation Columns



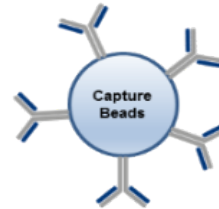
# MOST POPULAR ISOLATION TECHNIQUES



Differential  
Ultracentrifugation



Precipitation Solutions



ImmunoCapture



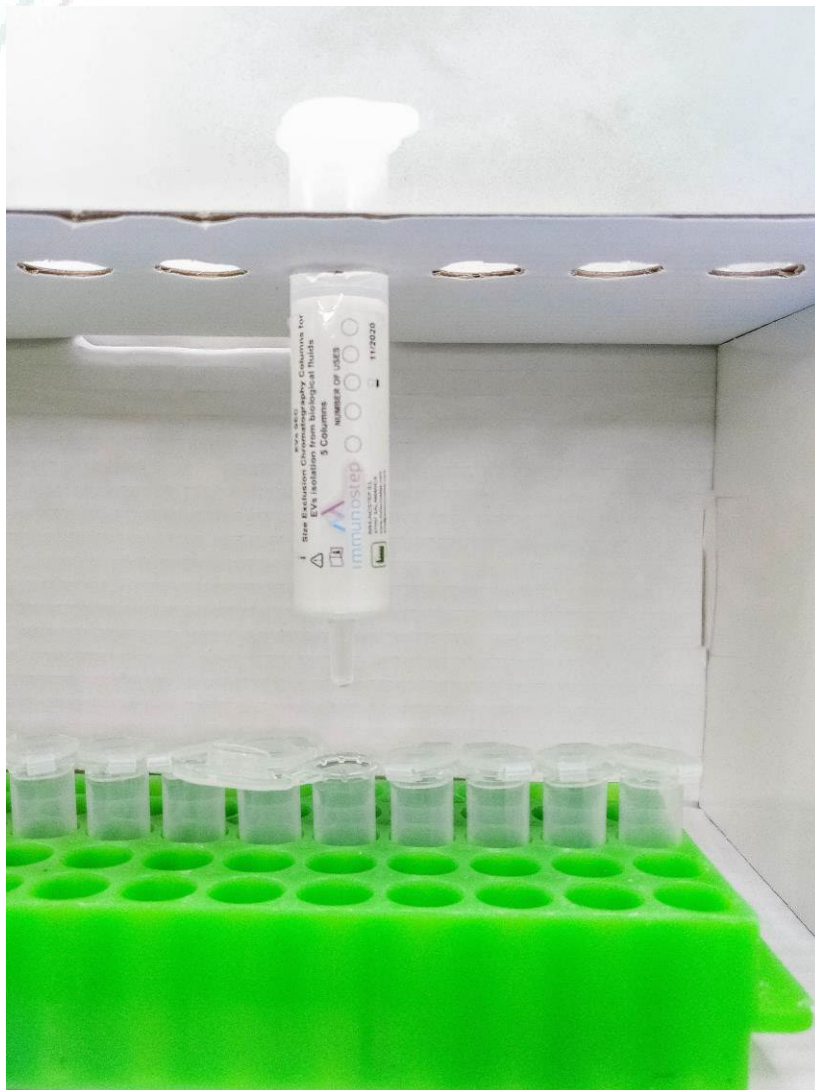
Size Exclusion  
Chromatography Columns

## SPECIFICATIONS

Separation size	70-1000nm
Volumetric Flow Rate at RT	<0.5 ml/min
Sample volume	<1 ml (500 µl optimal volume)
Column volume	10 ml
Void volume	3 ml
pH stability working range	3-13
pH stability cleaning in place (CIP)	2-14
Shelf life	12 months at 2-8°C

- ✓ Indicated for low volumes
- ✓ Rapid & Reliable Exosome Purification
- ✓ Standardisable & Reproducible
- ✓ Pure & Clean Isolated Samples

# Size Exclusion Chromatography Columns



## Preparation for use

1. Place the column in a holder and level it, make sure the column is vertical.
2. Remove the bottom cap.
3. Leave the top-cap carefully.

## Column equilibration

1. Rinse the column with 7 mL of elution buffer (PBS with sodium citrate).
2. Make sure enough time is given for the column to be in the operational temperature range.
3. Do not allow the column to run dry. The top filter must stay wet.
4. Use only fresh filtered (0.2  $\mu$ L) buffer to avoid particulate contamination.

## Sample fraction collection

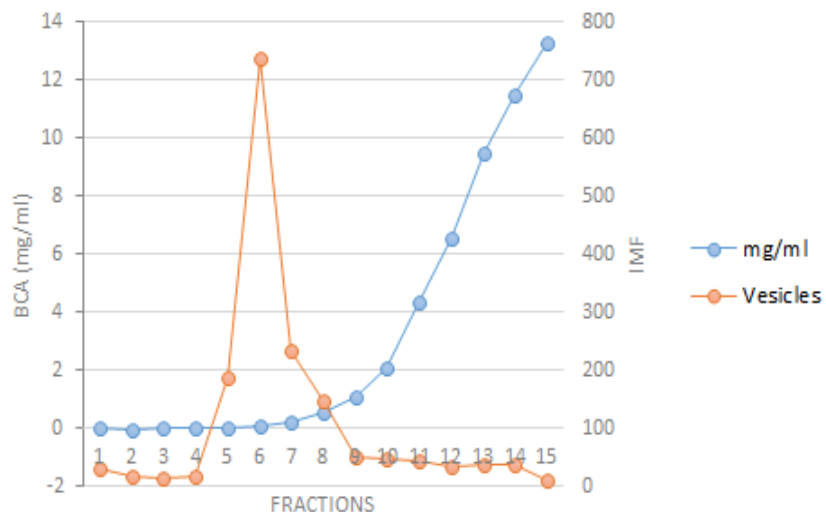
1. When the column is used according to the protocol, for 500 $\mu$ L fractions, the first four fractions (2.0 mL), is the void volume which does not contain vesicles, and they elute predominantly in fractions 5, 6 and 7 with removal of protein contamination. Fractions beyond 8 usually contain higher protein and lower vesicle levels (Fig 1).

## Post collection of vesicles

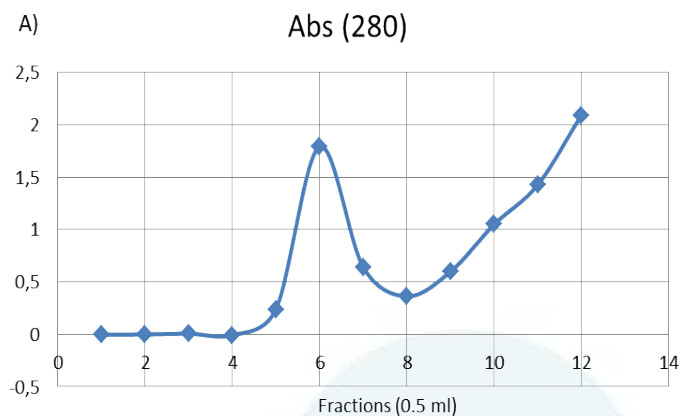
1. After the collection of the vesicle fractions, flush the column with 14 to 21 mL of equilibration buffer to flush out all the protein and small molecules before the next sample application. After that, flush the column with 7 mL of a bacteriostatic agent and store as indicated in the storage section

# SEC columns Performance

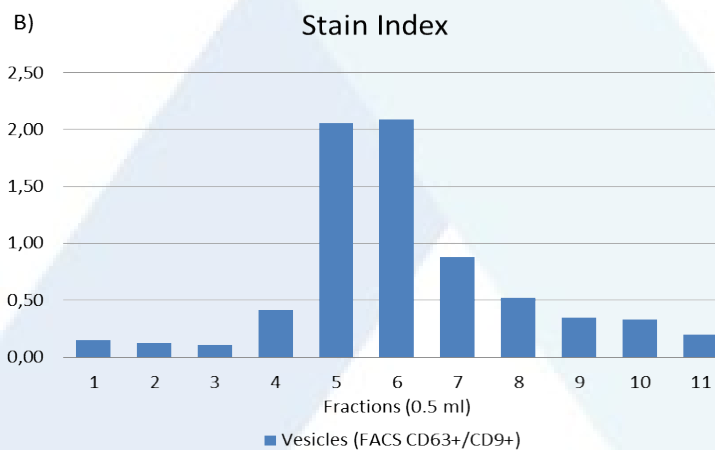
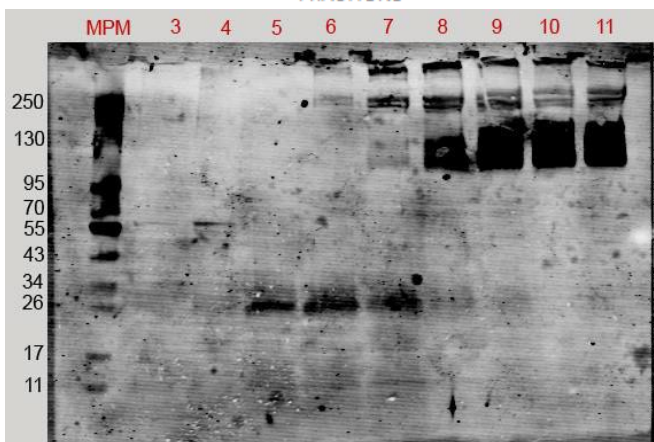
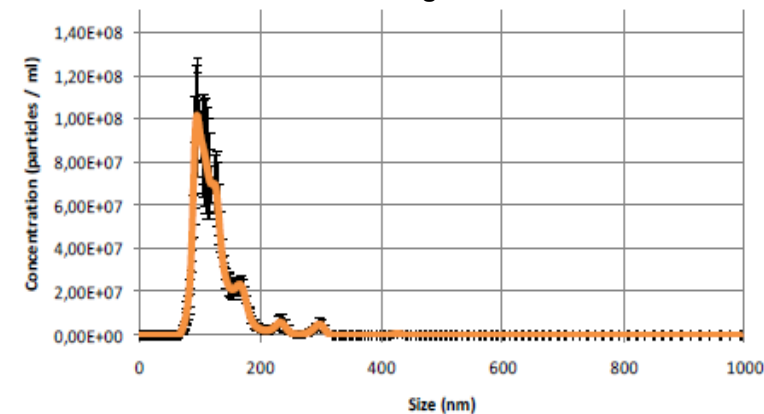
Elution profile



Exosome Plasma isolated by SEC

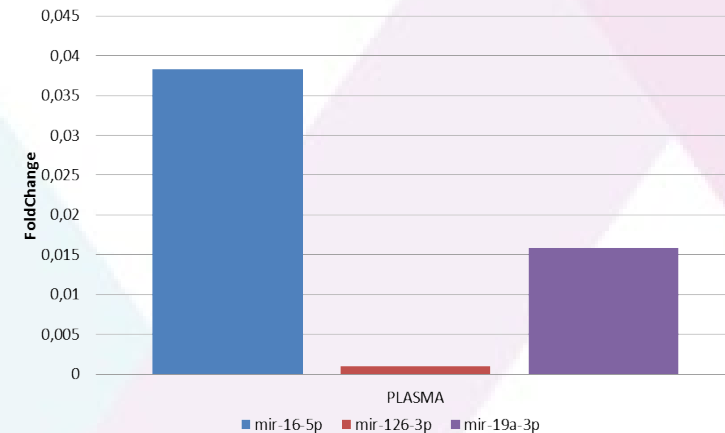


NanoSight



Stain index: (MFI positive- MFI background)/ 2σ background

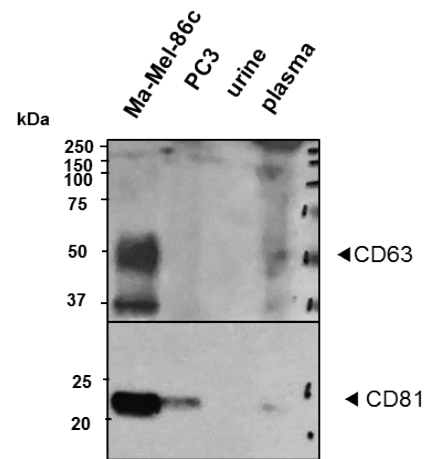
miRNAS



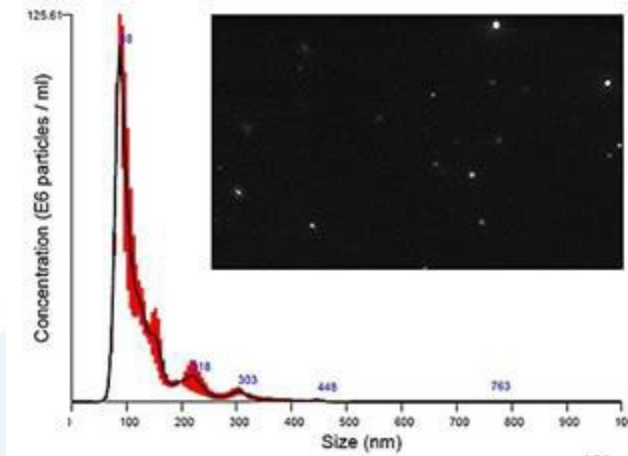
Exosomal miRNA levels by quantitative RT-PCR

# Common Methods to Analyze EVs Proteins

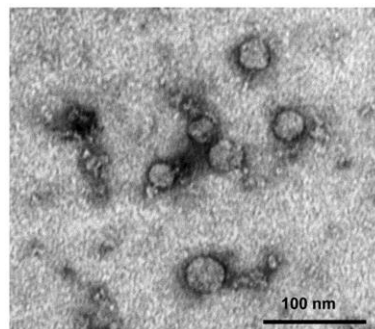
## Western Blot



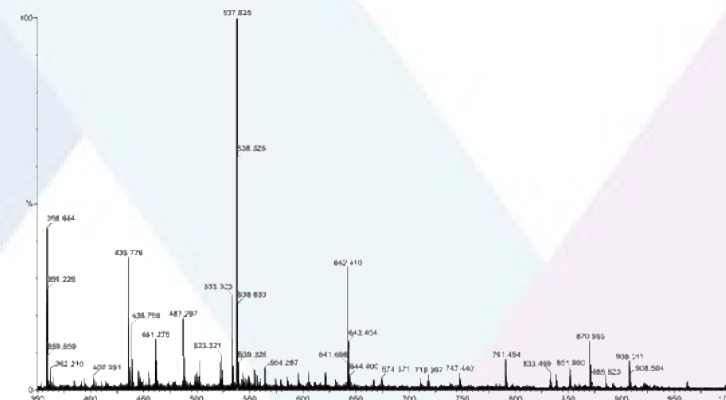
## Nanoparticle Tracking Analysis



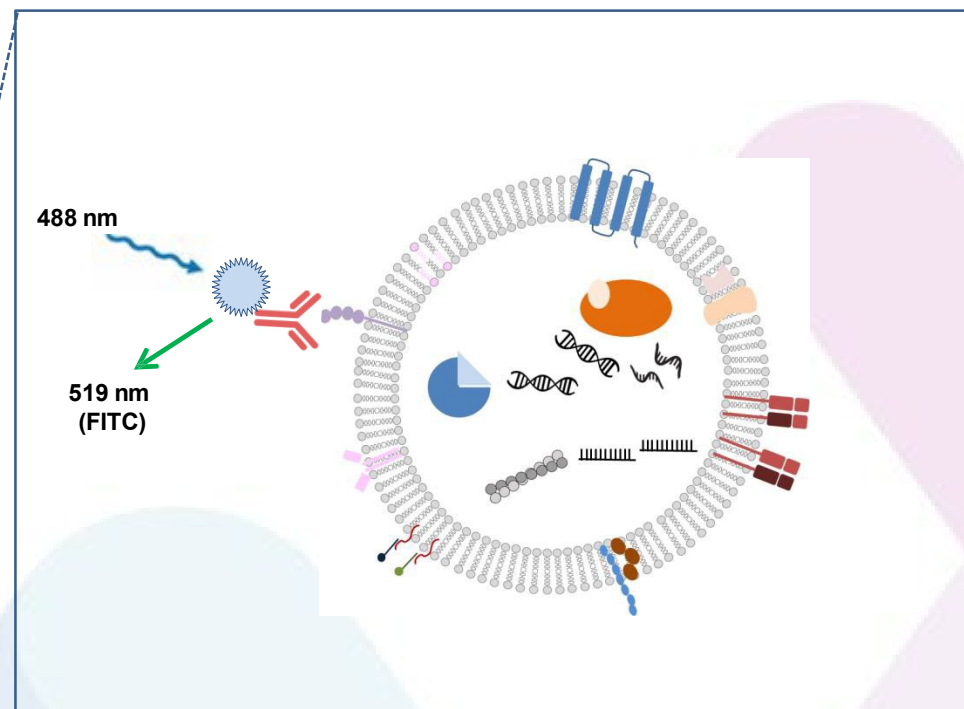
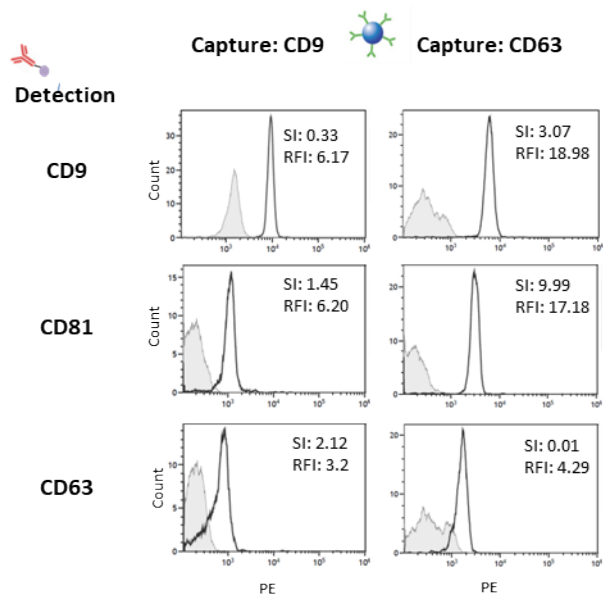
## Electron microscopy



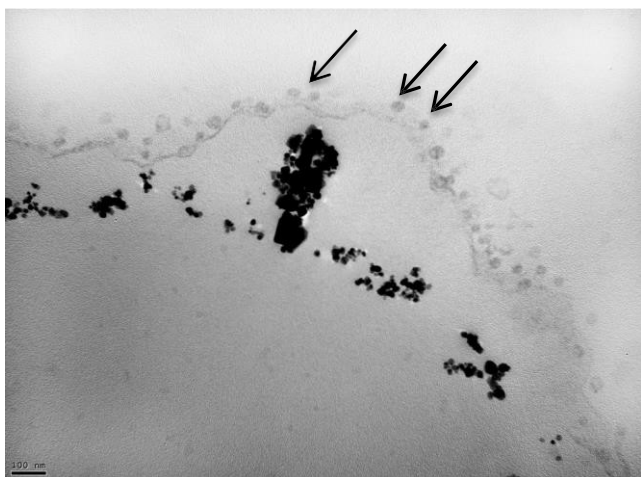
## Mass spectrometry



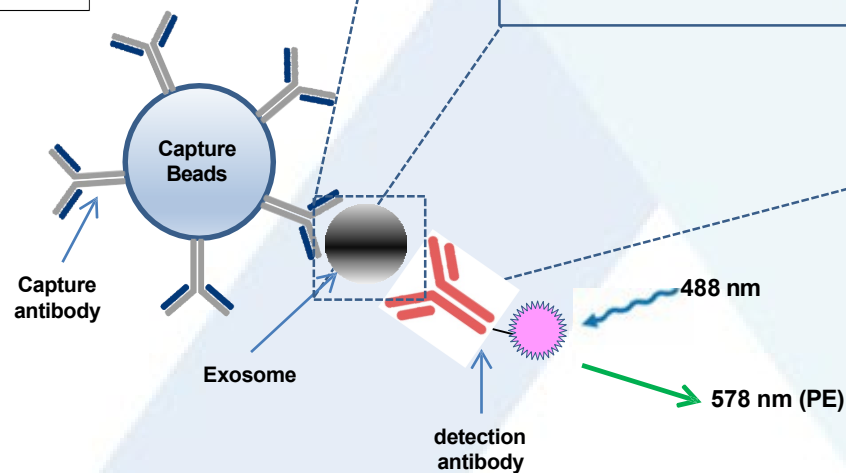
# ExoStep: bead-based principle



## Electron Microscopy



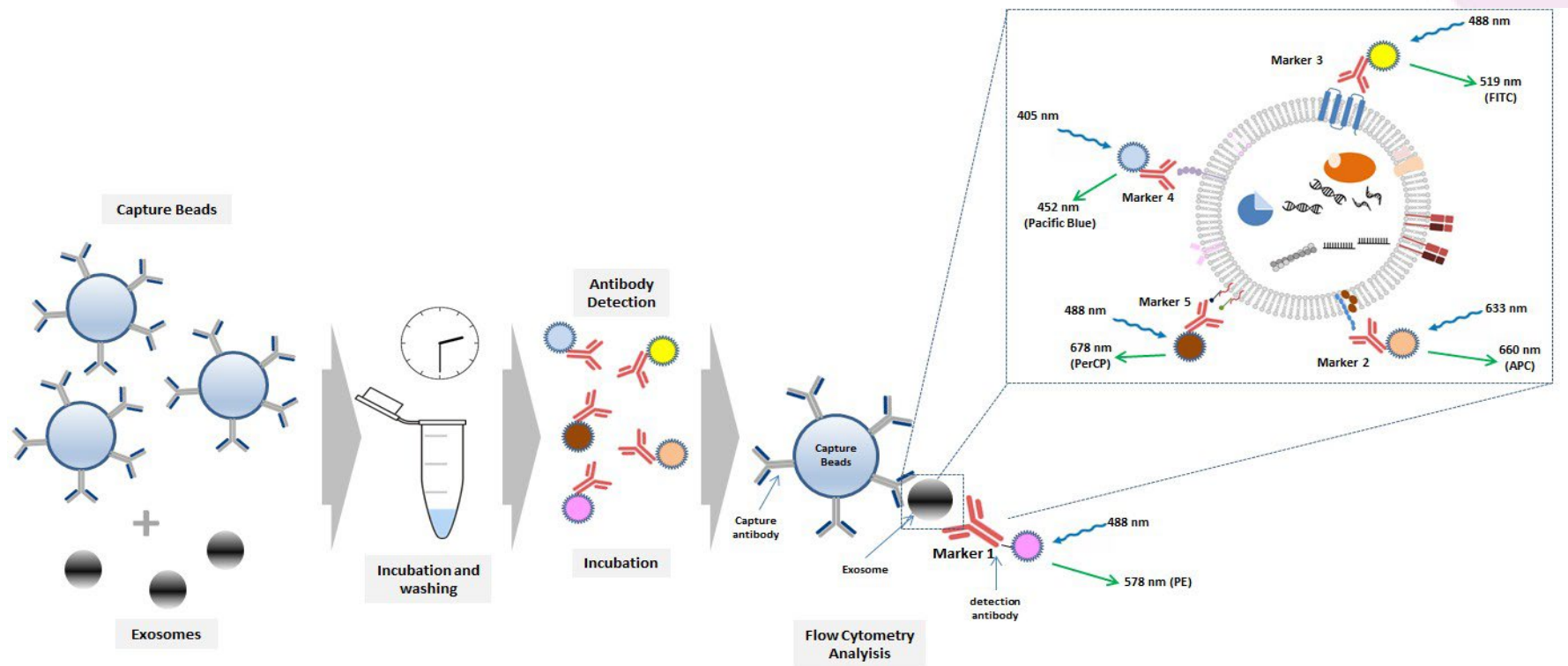
■ No EV  
□ 10<sup>9</sup> EV



# ExoStep Second Generation

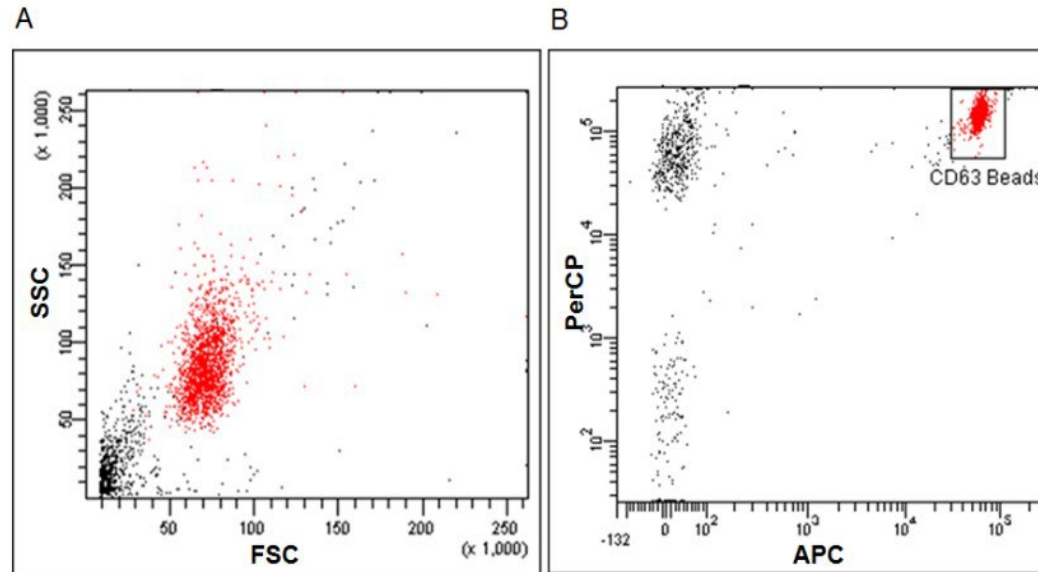
## Kit components:

- CD63+ (Clone TEA3/18) capture beads.
- Primary detection antibody, Anti-CD9 PE (Clone VJ1/20) or Anti-CD81 PE (Clone MEM38)
- Assay Buffer 10X, PBS 10% BSA, pH 7,4



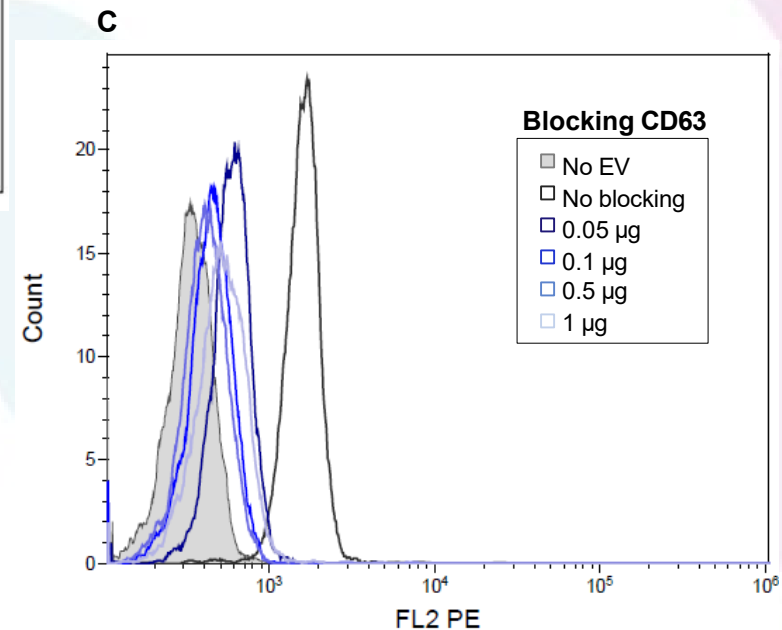


# Specific and unambiguous exosome detection by Flow Cytometry



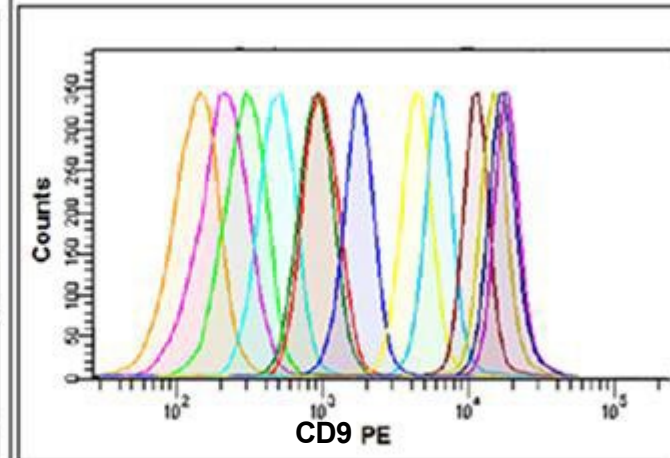
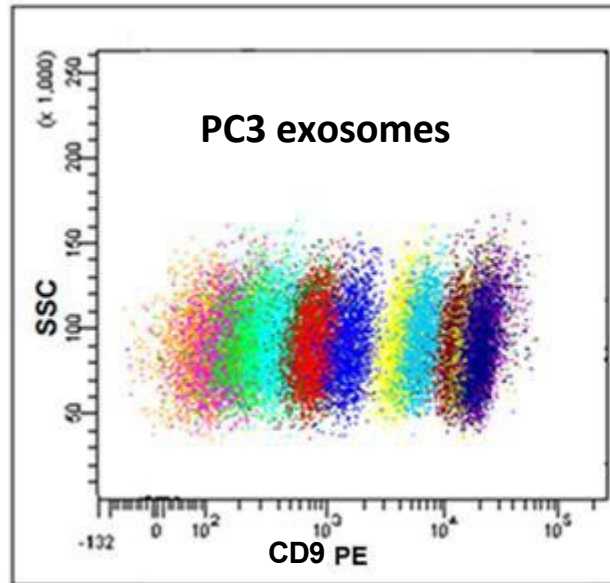
(A) Scatter Dot-plot of beads, FSC vs SSC. (B) Gating strategy on FL3 vs FL4 for flow cytometry acquisition and analysis, in order to remove doublets. Low-speed Flow Cytometry acquisition is recommended

(C) Specificity: antibody blocking. Capture CD63/Detection CD9-biot. 109 particles of PC3-derived EVs were pre-incubated with increasing amounts of the indicated soluble blocking antibody [anti-CD63 (Clone TEA3/18)] before being incubated for capture on CD63-coated beads.



# Dynamic range and limit of detection

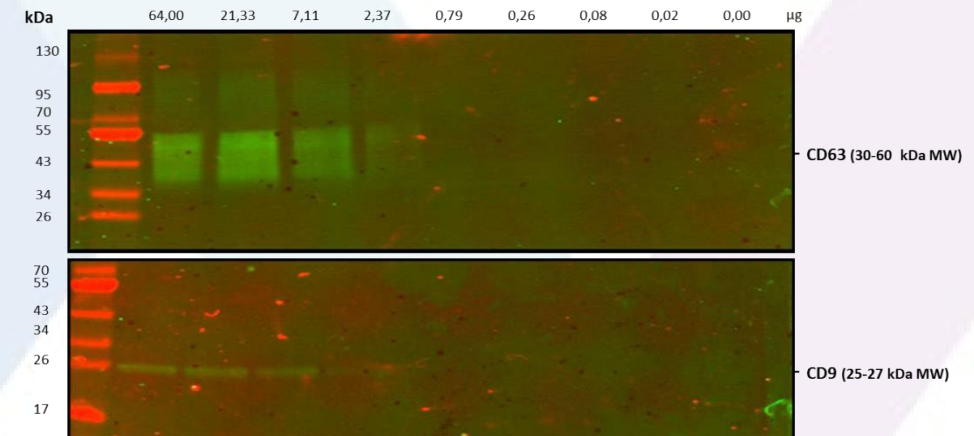
Greater sensitivity, wide dynamic range. Guaranteed detection even with small sample quantities



Sensitivity		
µg	MFI	SD*
0,0000	8,74	4,06
0,0625	14,20	8,49
0,1250	20,91	14,19
0,2500	36,85	52,61
0,5000	74,32	32,18
1,0000	79,86	82,30
2,0000	158,20	137,14
4,0000	380,00	248,41
8,0000	661,17	257,49
16,0000	1286,40	342,05
32,0000	1746,58	519,01
64,0000	2206,73	537,59
128,0000	2035,14	593,98

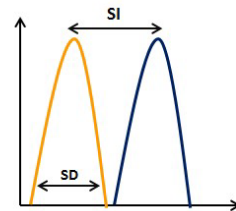
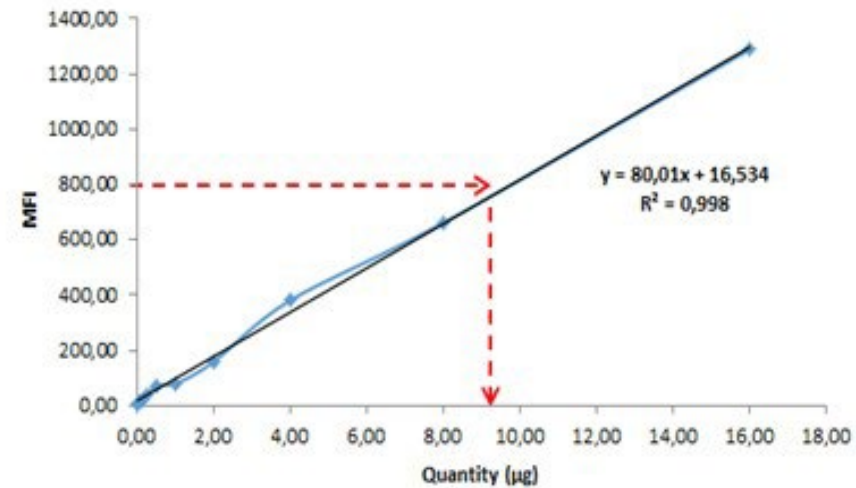
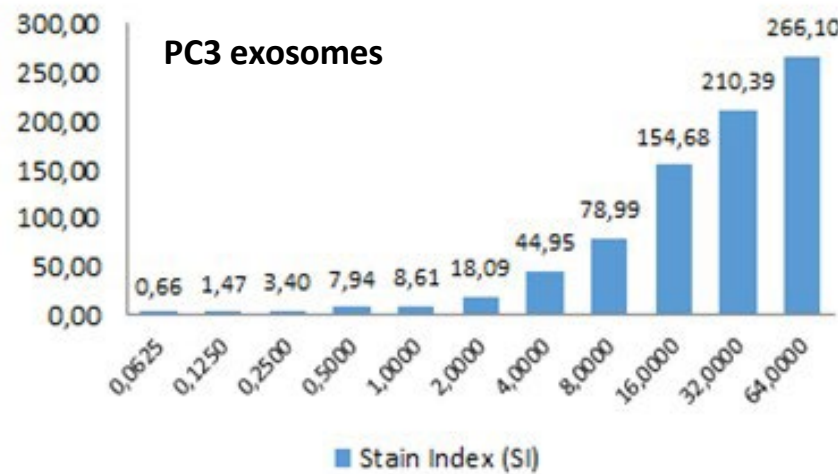
\* Standard Desviation.

- ✓ LOD: 0,0625 µg ->  $7,5 \times 10^7$  vesicles
- ✓ LOQ: 0,125 µg - Stain Index (SI) >1



# Quantitative analysis

Excellent correlation between fluorescence and the  
exosome quantity

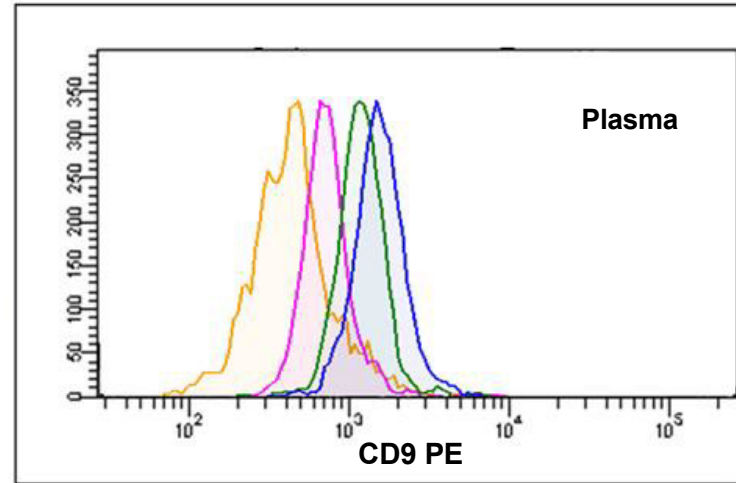
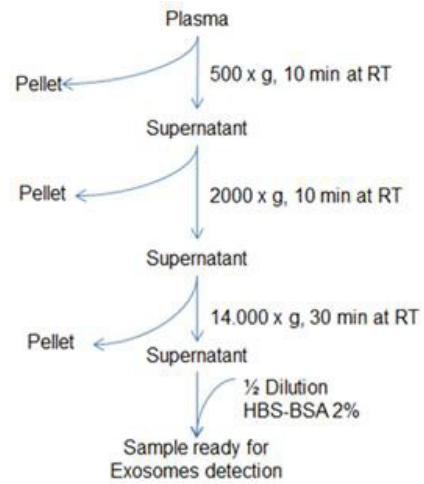


$$\text{Stain Index} = (\text{Median of Positive} - \text{Median of Negative}) / (\text{SD of Negative} * 2)$$



# Direct exosome detection in biological fluids

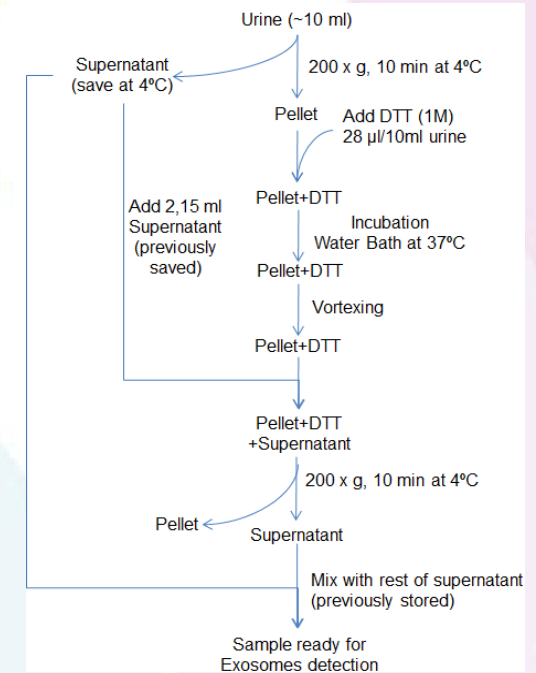
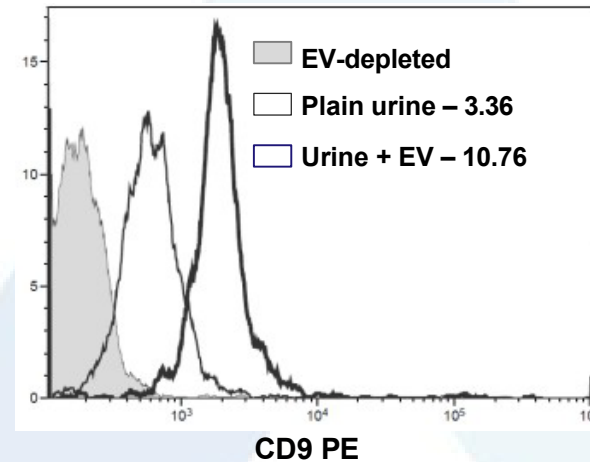
## Isolation no needed



**Sensibility for Direct Detection on Plasma (SI)**

Vol (ul)	CD63 Beads + CD9 biot + Strep PE
0,00	-
25,00	0,48
50,00	1,52
100,00	2,45

Stain Index (SI) = (Median of Positive - Median of Negative) / (SD of Negative \* 2)



Cell Culture

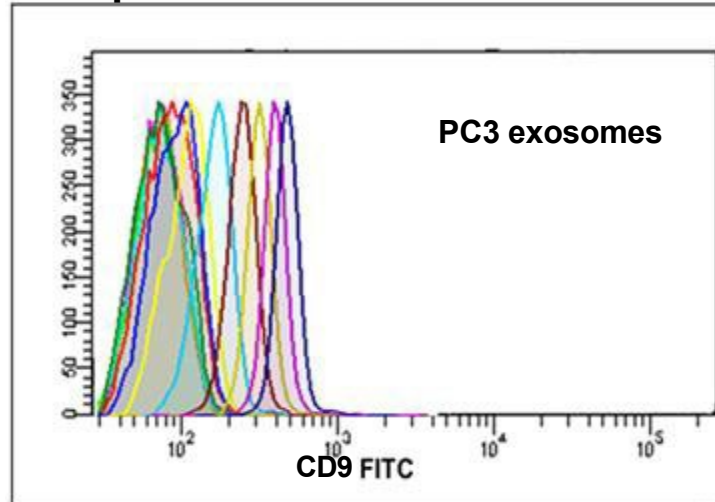
Plasma

Urine

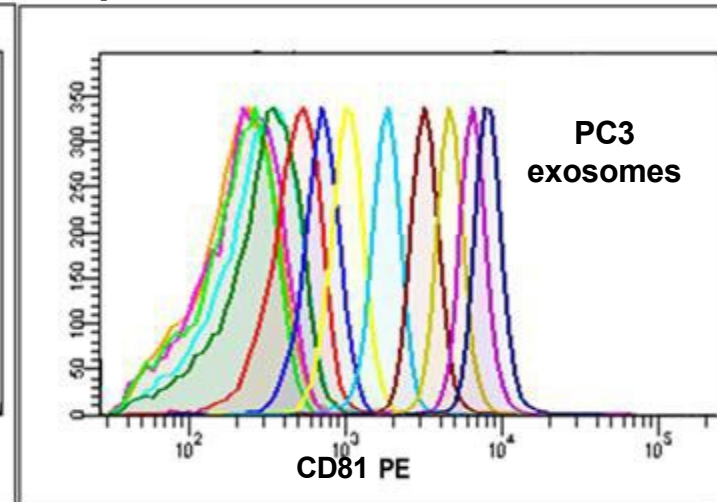
# Simultaneous Immunophenotyping

## Sensitivity detection in a wide range of quantities

Capture: CD63/ Detection CD9



Capture: CD63/ Detection CD81

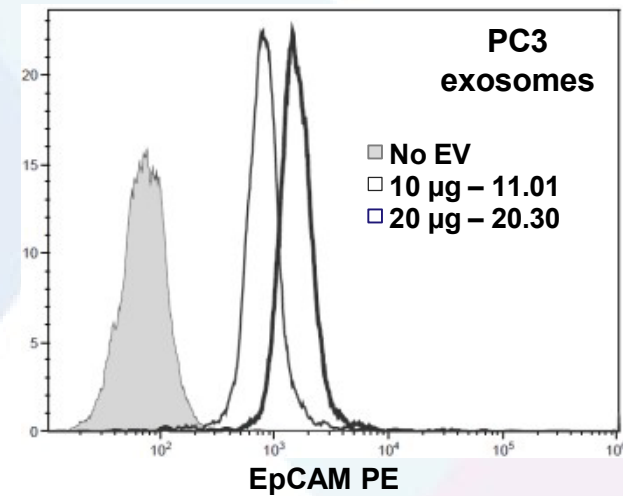


Immunophenotyping Sensitivity (SI\*)

µg	CD9-FITC	CD81- biot + Strep PE
0,0000	-	-
0,0625	-0,24	0,02
0,1250	-0,24	-0,04
0,2500	-0,30	0,30
0,5000	-0,24	0,45
1,0000	0,12	1,30
2,0000	0,24	2,46
4,0000	0,73	4,39
8,0000	2,06	9,65
16,0000	4,30	20,46
32,0000	6,54	33,62
64,0000	9,56	51,72
128,0000	13,77	69,17

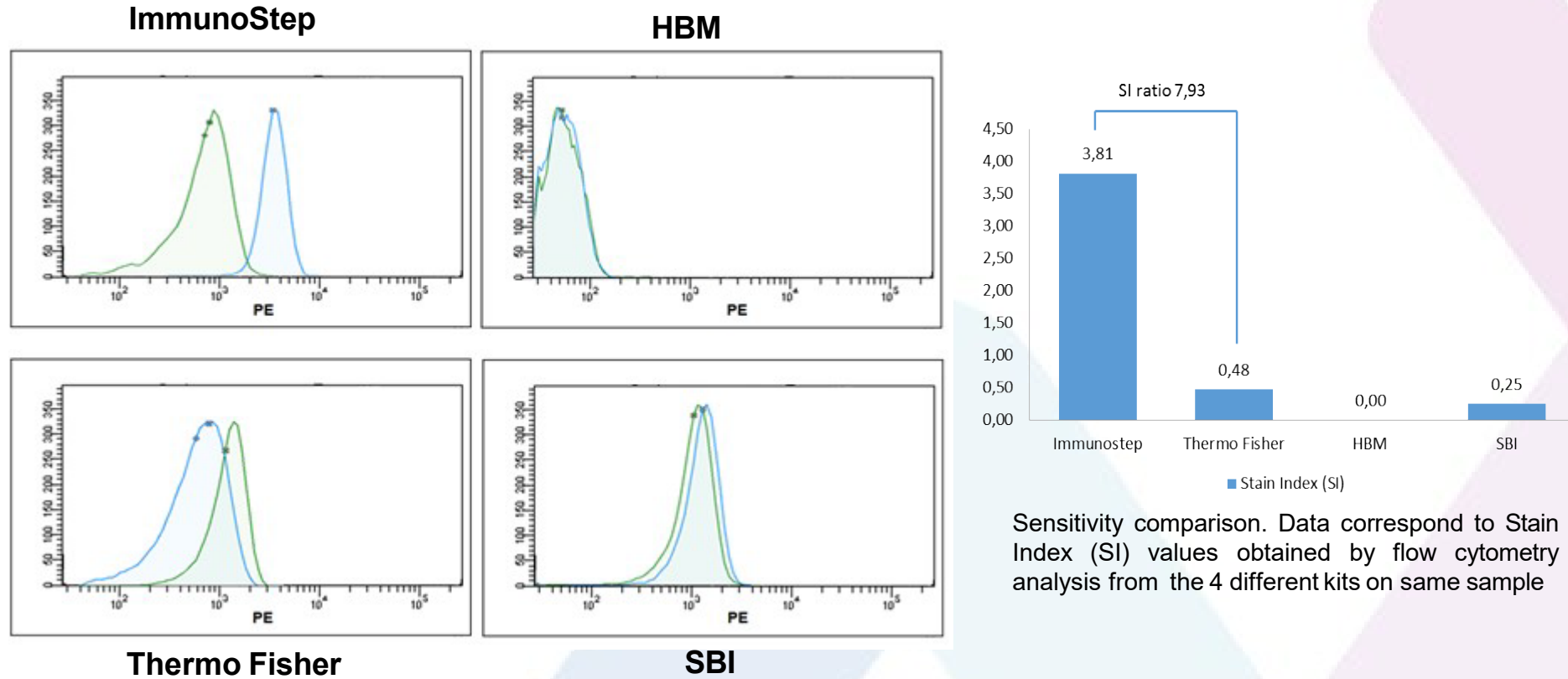
\*Stain Index (SI) = (Median of Positive - Median of Negative) / (SD of Negative \* 2)

Capture: CD63/ Detection EpCAM



# Competence and Competitive Advantage

## Sensitivity comparison on same sample



Sensitivity comparison. Data correspond to Stain Index (SI) values obtained by flow cytometry analysis from the 4 different kits on same sample

✓ **2 $\mu$ g - 2,40x10<sup>9</sup> PC3 exosomes**

# Exosome Analysis Summary

**ExoStep kit is a superior alternative for the sensitive detection of exosomes compared to the most commonly used methods, besides being easy to implement and analyze for any laboratory that has access to a conventional flow cytometer**

- Specific and unambiguous exosome detection.
- Quantitative analysis, excellent relationship between MFI and exosomes quantity.
- Direct detection of Exosomes in cell culture supernatant and biological fluids. Without isolation or precipitation.
- Very Small amount of sample needed
- Greater sensitivity, wide dynamic range. Guaranteed detection even with small sample quantities
- Reproducible
- Allowing simultaneous immunophenotyping of exosomes capture population



Thanks you for your attention

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